

The Metabolic Effects of Conjugated Linoleic Acids (CLA) in Chickens: A Review

Review Article

M. Royan^{1*} and B. Navidshad²¹ Department of Animal Biotechnology, Agricultural Biotechnology Research Institute (ABRI), Rasht, Iran² Department of Animal Science, University of Mohaghegh Ardabili, Ardabil, Iran

Received on: 27 Apr 2014

Revised on: 20 Jun 2014

Accepted on: 30 Jun 2014

Online Published on: Sep 2015

*Correspondence E-mail: m.royan@abrii.ac.ir

© 2010 Copyright by Islamic Azad University, Rasht Branch, Rasht, Iran

Online version is available on: www.ijas.ir

ABSTRACT

Conjugated linoleic acids (CLA) are natural constituents of meat and dairy products from ruminants, originating from bacterial biohydrogenation in the rumen. CLA supplementation increases the health benefits of animal-derived foods. There are inconsistent reports of the effects of dietary CLA on chicken's performance; however, the majority of previous reports cite anti-lipogenic effects of CLA. Diets could be formulated to increase n-3 fatty acid (FA) concentration in chicken meat by feeding n-3 FAs as a replacement for n-6 FA rich ingredients. Off-flavors and the chances of oxidative deterioration during storage of meat have been attributed to high n-3 FA levels in poultry diets. An approach to increase the n-3 FA content and decrease the n-6/n-3 FAs ratio in meat, using conjugated linoleic acid in diets, has been suggested. This review deals with the main topics of CLA metabolic effects in chickens.

KEY WORDS chickens, conjugated linoleic acid, gene expression, lipid metabolism.

INTRODUCTION

In 1979, a research group from the University of Wisconsin-Madison detected a compound with anti-mutagenic activity in ground beef extract (Pariza and Hargraves, 1985); because of its structural similarity to linoleic acid, it was named conjugated linoleic acid or CLA (Pariza *et al.* 1979; Ha *et al.* 1987). Later, it was found in many other food sources, especially in dairy products and in animals that have a ruminant-like, fermentative digestive process, alike wallabies and kangaroos (Chin *et al.* 1992; Joo *et al.* 2002). In the rumen, CLA is primarily synthesized by bacterial biohydrogenation of linoleic acid (18:2n-6) by a variety of bacterial species including *Butyrivibrio fibrosolvens* (Kepler *et al.* 1966; Griinari *et al.* 2000). CLAs are derived mainly from the microbial activity of the rumen. Studies have found that cis-9, trans-11 CLA can also be endogenously synthesized via $\Delta 9$ desaturation of trans-vaccenic acid in the mammary glands of cows, and monogas-

trics such as mice and humans (Adolf *et al.* 2000; Santora *et al.* 2000; Turpeinen *et al.* 2002; Mosley *et al.* 2006). It was been reported that the endogenous synthesis of CLA is the most important source of the cis-9, trans-11 CLA isomer in milk fat, representing up to 64-78% of the total CLA (Corl *et al.* 2001; Griinari and Bauman, 1999). Based on the high correlations between CLA and trans-vaccenic acid, it was suggested that desaturation of vaccenic acid might be the main source of CLA in muscle lipids (Knight *et al.* 2003). In the liver of rats, delta 11-trans octadecenoic acid is enzymatically converted to the delta 9-cis, delta 11-trans octadecadienoic acid (Banni and Martin, 1998). It has been reported that the bacterial population of rodent intestine may also converted unsaturated fatty acids to CLA (Chin *et al.* 1994).

Differential effects of CLA isomers

CLAs have a variety of biological activities. CLA in chicken nutrition One reason for the diversity of biological effects of CLA is

that CLA is a blend of geometric and positional isomers, with double bonds located at the Δ [9,11], [10,12], [8,10], [7,9] and [11,13] positions. Although several CLA isomers exist in food (Kramer *et al.* 1998), most research is focused on the two major isomers, cis-9, trans-11 and trans-10, cis-12. Anti-cancer activity which has been shown for both major isomers of CLA can be additive (Ip *et al.* 2002; Masso-Welch *et al.* 2002; Masso-Welch *et al.* 2004). Moreover, it was observed that cis-9, trans-11 and trans-10, cis-12-CLA may affect lipid metabolism in diverse tissues, including liver, muscle and adipose tissue, in different ways (Evans *et al.* 2002).

These two isomers can work independently. The cis-9, trans-11 isomer improves growth performance in rodents. However, body fat reduction, inhibition of stearoyl-CoA desaturase and reduction of hepatic apolipoprotein B secretion resulted exclusively from trans-10, cis-12 CLA isomer activity (Chin *et al.* 1994; Cook *et al.* 1993; Park *et al.* 1999a; Pariza *et al.* 2000; Storkson *et al.* 2005; Valeille *et al.* 2004). These two isomers have antagonistic effects on each other. The cis-9, trans-11 isomer has a potential anti-diabetogenic effect and can correct insulin resistance while the trans-10, cis-12 isomer promotes insulin resistance (Song *et al.* 2004).

CLA and human health

The increased interest in the daily intake of CLA in humans results from evidences issued from animal models, which indicates the potential health benefits of CLA (Williams, 2000). CLA has been reported to act as a fat-to-lean repartitioning agent (Park *et al.* 1997), as a growth modulator (Chin *et al.* 1994) and hypocholesterolemic and antiatherogenic agent (Lee *et al.* 1998). In animal models, CLA has been shown to reduce the incidence of skin, for estomach, colon, mammary, and liver cancers (Bhattacharya *et al.* 2006; Kelley *et al.* 2007; Lee *et al.* 2005). Milk fat, being the richest natural source of CLA, has a protective effect against the incidence of human breast cancer (Knekt *et al.* 1996).

Larsson *et al.* (2005) reported a negative correlation between CLA consumption and colorectal cancer incidence in a 15-year study. Furthermore, CLA have beneficial effects on lowering blood cholesterol (Bhattacharya *et al.* 2006; Kritchevsky, 2000; McLeod *et al.* 2004).

CLA in animal tissues

In animals, the highest CLA concentration was reported in the adipose tissue of kangaroos (38 mg/g fatty acids) (Engelke *et al.* 2004). In farm animal products, foods originating from ruminants are the main source of CLA, with the cis-9, trans-11 (rumenic acid) and trans-10, cis-12 isomers comprising 80-90% and 3-5% of the total CLA, respectively (Khanal and Dhiman, 2004).

Non-ruminant meats contain lower concentrations of CLA. For example, chicken and pork contain 0.9 and 0.6 mg/g, re-

spectively (Chin *et al.* 1992). Level of CLA differs between fat meat and lean meat. Fat meat has a much higher concentration of CLA (cis-9, trans-11: 960-1310 mg/100 g) than lean meat (cis-9, trans-11: 6-43 mg/100 g) (Fogerty *et al.* 1988). CLA content in non-ruminants body fat is considerably lower than in the ruminants fat (Fogerty *et al.* 1988). Fritsche and Steinhart (1998) found no variation in the CLA levels or isomer distribution in bulls and bullocks fat samples and suggested that the CLA content of meat is independent of the hormonal status. Fish oils also contain low CLA concentrations (Chin *et al.* 1992). CLA is not found in any of the common plant oils. Small amounts of CLA maybe formed during their heating, bleaching and deodorisation in the refining process (Ip *et al.* 2002).

In vitro synthesis of CLA

The 50%-50% mixtures of CLA bioactive isomers are manufactured commercially through alkaline isomerisation or partial hydrogenation of either linoleic acid or, more frequently, sunflower or safflower oils that are rich in linoleic acid (Banni, 2002). Synthetically manufactured CLA includes equal levels of the cis-9, trans-11-CLA and trans-10, cis-12-CLA and significant levels of other CLA isomers (Pariza *et al.* 2001) in comparison to natural sources where the cis-9, trans-11-CLA isomer prevails.

CLA and fat metabolism

The trans-10, cis-12 isomer is responsible for fat reduction (Park and Pariza, 2007; Park *et al.* 1999a). The body fat-reducing effect of CLA is probably caused by CLA itself rather than its metabolites (Park and Pariza, 2007). The non-acidic derivatives of CLA were found not effective on fat metabolism, indicating that the carboxyl end is needed for CLAs activities (Cook *et al.* 2000; Park *et al.* 2004). *In vitro* experiments showed that CLAs stimulate modifications in the membrane of adipocytes and regulate its gene expression, resulting in a decrease in the activity of the delta-9 desaturase enzyme (Hur *et al.* 2007). The effects of CLA on body composition are mediated by the increase in energy expenditure, the lowering of body fat deposits, the stimulation of the apoptosis process in pre-adipocytes and an increase in lipolysis and β -oxidation in muscle tissue (Park and Pariza, 2007).

It was suggested that CLAs promote fatty acid β -oxidation in skeletal muscle (Pariza *et al.* 2001; Wahle *et al.* 2004). This effect is shown by an up-regulation of carnitine palmitoyltransferase I (CPT-I, the main enzyme for β -oxidation) in skeletal muscles (Bouthegourd *et al.* 2002; Degrace *et al.* 2004; Nagao *et al.* 2005; Park *et al.* 1997; Peters *et al.* 2001). CPT-I initiates the fatty acid transport into the mitochondrion for β -oxidation. In rat and mice adipose tissue, as well as in mice muscle, CLA up-regulate this enzyme (Park *et al.* 1997; Rahman *et al.* 2001). As a consequence, CLA increases the energy

supply from fat sources and helps to decrease body fat deposition. Higher peroxisomal β -oxidation activity has been reported in animals fed CLA (Moya-Camarena *et al.* 1999; Yamasaki *et al.* 2001; Choi *et al.* 2004; Degrace *et al.* 2004). A higher fatty acid β -oxidation may reduce the availability of fatty acids for triglyceride synthesis, thus reducing fat deposition (Mersmann, 2002).

It has been reported that CLA might decrease leptin secretion (Inoue *et al.* 2004). This effect maybe due to the lower total adipose tissue in CLA-treated subjects, although the increased food intake in result of reduced levels of leptin was not been reported in such studies. CLA enhanced adiponectin and reduced TNF- α (Inoue *et al.* 2004; Nagao *et al.* 2003; Nagao *et al.* 2005; Pariza *et al.* 2000), which may improve insulin sensitivity; consequently, it can become an important mediator in several chronic disorders including obesity (Hotamisligil and Spiegelman, 1994). Alteration of interleukins by CLA was also reported (Bassaganya-Riera *et al.* 2003; Changhua *et al.* 2005). In the study of Royan *et al.* (2011a), dietary CLA effectively decreased fat deposition in broiler chicken meat, especially in breast tissue. Similarly, Kawahara *et al.* (2009) report that the total lipid and triglyceride content in breast meat tended to decrease in broilers fed 1-2% dietary CLA. Moreover, in the study by Kawahara *et al.* (2009), dietary CLA reduced the amount of thiobarbituric acid reactive substances (TBARS) in raw chicken meat during storage at 4 °C for 5 days. These results provided evidence that CLA feeding can be a practical strategy not only for adding nutritional benefits to chicken meat but also to improve meat quality, including oxidative stability.

CLA and energy expenditure

CLA may decrease body fat by promoting energy expenditure (Park and Pariza, 2007). West *et al.* (1998) demonstrated that dietary supplementation with CLA enhanced the energy expenditure in animals. This increase in energetic expenditure is sufficient to explain the lower fat accumulation in CLA-supplemented mice (Atkinson, 1999). The CLA mediated increase in energy expenditure is accompanied by higher oxygen consumption (Choi *et al.* 2004; Ohnuki *et al.* 2001; Terpstra *et al.* 2002; West *et al.* 2000) and the up-regulation of uncoupling proteins (UCPs) (Choi *et al.* 2004; Nagao *et al.* 2003; Peters *et al.* 2001), which are both indicators for energy expenditure.

It seems that uncoupling protein 2 (UCP2) acts to disconnect energy conservation from substrate oxidation, leading to higher heat production and lower energy conservation (Mersmann, 2002). Dietary CLA increased UCP2 in white and brown adipose tissue of mice (Tsuboyama-Kasaoka *et al.* 2000); nevertheless, in another report, CLA increased this protein only in brown, but not in white, adipose tissue (West *et al.* 2000). A lower respiratory quotient (RQ) is an indicator of increased fat oxidation. A reduced RQ was reported in mice

fed a low-fat diet containing CLA, but not with inclusion of CLA in a high-fat diet (West *et al.* 1998). This effect of CLA might be species specific, because in pigs and sows fed CLA, the RQ was not affected (Muller *et al.* 1999; Muller *et al.* 2000).

CLA and adipose cells

CLA reduces adipose cell mass and number. This effect is attributed to inhibiting lipoprotein lipase (LPL) and stearoyl-CoA desaturase activities in adipose cells, increasing apoptosis of preadipocytes and adipocytes and modulating lipolysis. Lipoprotein lipase is the main enzyme for fat absorption and its inhibition in adipocytes decreases fat uptake (Park *et al.* 1997; Park *et al.* 1999b; Park *et al.* 2004). This inhibitory effect has been linked to the trans-10, cis-12 isomer but not cis-9, trans-11 (Lin *et al.* 2001; Park *et al.* 1999b; Park *et al.* 2004). LPL is synthesized in adipose tissue and transferred to the endothelial cell surface, where it functions to remove fatty acids from blood lipoproteins. The released fatty acids can then move into the adipose tissue to be oxidized or to supply building blocks for the synthesis of complex lipids like triglycerides (Mersmann, 2002). A lower LPL activity would reduce the fatty acid available for triglyceride synthesis, and consequently would decrease lipid accumulation. Although it was been reported that very low levels of CLA increased LPL activity in 3T3-L1 cells, higher CLA levels inhibited the enzyme activity (Park *et al.* 1997; Park *et al.* 1999b; Lin *et al.* 2001).

The role of stearoyl-CoA desaturase (known also as SCD-1 or $\Delta 9$ -desaturase) is the $\Delta 9$ -cis desaturation of many fatty acids including palmitoyl and stearoyl-CoA. Desaturation of these SFA is essential to generate mono unsaturated fatty acids (MUFA) necessary for inclusion into the sn-2 position of triglycerides (Cohen *et al.* 2002). It was shown that CLA consumption could down-regulate stearoyl-CoA desaturase (Hur *et al.* 2007). Therefore, CLA mediated inhibition of stearoyl-CoA desaturase activity may reduce body fat mass (Choi *et al.* 2002). The higher SFA / MUFA ratios in body fat following CLA feeding suggests that down-regulation of SCD-1 may be one explanation for CLA's lipid-lowering effects (Choi *et al.* 2000). Thus, CLA anti-lipogenic effects are mediated by inhibiting lipogenesis and triglyceride esterification by an interruption in the fatty acid desaturation process. Bretilon *et al.* (1999) reported that $\Delta 6$ desaturation of linoleic acid was reduced when the dietary ratio of cis-9, trans-11 CLA to linoleic acid increased, but the trans-10, cis-12 CLA isomer had only a small effect on the desaturation process and was only effective only at the highest levels. They also suggested that $\Delta 9$ -desaturase activity is repressed only by the trans-10, cis-12 CLA isomer. Another direct effect of CLA on lipid metabolism respect the decrease in the adipose tissue deposit by enhancing apoptosis of preadipocytes and adipocytes (Brodie *et al.* 1999; Brown *et al.* 2001; Brown *et al.* 2003; Cohen *et al.*

2002). Some reports indicate that the fat-reducing effects of the trans-10, cis-12 CLA isomer are not due to increased apoptosis (Xu *et al.* 2003) but are probably due to a reduction in preadipocyte differentiation into mature adipocytes (Brodie *et al.* 1999; Satory and Smith, 1999; Evans *et al.* 2000; Simon *et al.* 2005).

CLA and lipolysis

CLA's effect on adipocytes may be mediated through the peroxisome proliferator-activated receptor-gamma (PPAR γ). PPAR γ is a member of the nuclear receptor superfamily and regulates the differentiation, proliferation and lipogenesis processes in adipocytes (Gregoire *et al.* 1998; Tontonoz *et al.* 1994). Down-regulation of PPAR γ could lead to the effects caused by CLA. CLA has been reported to decrease PPAR γ expression (Brown *et al.* 2001; Brown *et al.* 2003; Choi *et al.* 2000). It has also been shown that CLA strongly down-regulates PPAR α , the predominant PPAR form in liver, with an antilipogenic effect (Moya-Camarena *et al.* 1999). Royan *et al.* (2011b) showed that the adipose PPAR γ gene expression in palm oil-fed birds was significantly up-regulated; however, there were no significant differences in PPAR γ gene expression in the adipose tissue between birds fed diets containing CLA, fish oil, soybean oil, or the mixture of these fats.

CLA and inhibition of adipocyte lipid synthesis

Accretion of triglycerides in adipocytes is the main indication for the growth of adipose tissue; therefore, a reduction in adipocyte lipid synthesis would reduce fat deposition (Mersmann, 2002). The effect of CLA on decreasing adipocyte hypertrophy was reported in the mouse (Tsuboyama-Kasaoka *et al.* 2000) and rat (Azain *et al.* 2000). Therefore, a combination of lower glucose and fatty acid uptake and reduced de novo fatty acid and triglyceride synthesis, is expected to be the reason for the lower fat accumulation in the adipocytes of animals fed CLA. Because CLA is not as effective for reducing hepatic lipogenesis (Choi *et al.* 2000), it was suggested that CLA lacks significant antilipogenic effects in birds (Du and Ahn, 2002).

Interaction of CLA with other fatty acids

Research examining the interaction between dietary CLA and fat level showed that CLA concentrations of 0.5-1% reduced fat mass equivalently in mice fed either a high-fat (45% of calories) or a low-fat (15% of calories) ratio (Delany *et al.* 1999). Therefore, it seems possible that the reduction of body fat through CLA supplementation is independent of dietary fat intake, at least in mice (Kennedy, 2007). It is obvious that the nature of dietary lipid used does affect the anti-adipogenic capability of CLA (Kennedy, 2007). Both the CLA and polyunsaturated fatty acids (PUFAs) are lipid metabolism modifiers; consequently, the use of CLA in combination with different PUFAs in the diet may improve the productive efficiency

and reduce body fat deposition (Zanini *et al.* 2006). When CLA was combined with oleic, linoleic, or linolenic acids, the negative effects of higher saturated fatty acids were reduced (Kim, 2007).

Royan *et al.* (2013) reported that dietary CLA, either alone or in combination with soybean oil or fish oil significantly increased the CLA content of breast and thigh tissues as compared to CLA-free diets; however, the CLA content of tissues did not exhibit a dose-dependent response to CLA supplementation. In both the breast and thigh tissues, the combination of CLA and soybean oil resulted in more CLA deposition than the CLA or CLA combined with fish oil-containing diets. Herzallah (2013) demonstrated that lactic acid bacteria of animal origin (*L. reuteri*) significantly enhanced CLA synthesis in both eggs and broiler meat cuts.

Zanini *et al.* (2008) showed that a dietary mixture of CLA and soybean oil increased total deposited fatty acids, including CLA, in meat, due to the higher lipid content in meat. The dietary CLA and canola oil mixture reduced SFA and MUFA and increased CLA content, with lower total lipid content in meat.

Zanini *et al.* (2006) reported that the fat-reducing effect of the mixture of CLA and canola oil was only observed in female birds, which normally have higher lipid accumulation than males. Moreover, they observed a lower total lipid content of the gizzard and a lower relative liver weight in females fed the CLA and canola oil mixture. However, when CLA in association with soybean oil was used, the total lipid content of the liver and gizzard increased linearly.

Aydin *et al.* (2001) reported that the use of CLA in association with oils rich in n-3 fatty acids optimized the CLA effect. In another study, the CLA in combination with soybean oil or coconut decreased body mass and epididymal fat mass in mice (Kennedy, 2007). However, CLA and fish oil in combination showed no effect on adiposity (Hargrave *et al.* 2005). Mixtures of CLA and n-3 PUFA accelerated recovery by activating PPAR δ (i.e., induced UCP3) and upregulating the expression of KGF in the colon (Bassaganya-Riera and Hontecillas, 2006). Brown *et al.* (2001) have also shown that the CLA effect can be modified by the oil supplement; therefore, the association of CLA with other fats should be considered when interpreting the results of feeding studies using CLA in combination with other fats. The effect of CLA in reducing total serum cholesterol, liver weight and total lipid content of giblets of broiler chickens was also dependent on the oil source (Zanini *et al.* 2006). Shin *et al.* (2011) studied the effects of the combination of dietary CLA and n-3 fatty acids on the linoleic acid (C18:2n-6) and arachidonic acid (C20:4n-6) concentrations of broiler chicken breast and thigh muscles, and found that the combination of CLA and menhaden fish oil is effective to reduce the concentrations of linoleic and arachidonic acids in broiler chicken breast and thigh muscles.

Effects of CLA on PPARs expression

The anti-lipogenic effect of CLA is mediated through reducing uptake and transport of fatty acids as evidenced by lower lipoprotein lipase (LPL) and fatty acid binding protein (FABP) levels (Park *et al.* 1999a). It is suggested that for decreasing lipogenesis the trans-10, cis-12 isomer is a potent anti-lipogenic agent (Peterson *et al.* 2003). It seems that CLA applies the anti-lipogenic effects at the transcriptional level by regulating gene expression of important regulatory proteins and enzymes to some extent by PPARs (Berge *et al.* 2004).

Some studies showed that CLA could act as high-affinity ligands for PPARs isotypes, mainly PPAR α and could alter the level and change the activation of different PPARs (Moya-Camarena *et al.* 1999). The majority of previous reports, refer that the trans-10, cis-12 and the cis-9, trans-11 isomers of CLA act as activator ligands of PPAR α and δ , but activation of PPAR γ was small (Houseknecht *et al.* 1998; Martin *et al.* 2000; Evans *et al.* 2002). The PPAR γ mediated regulation of LPL and FABP support the idea that the antilipogenic effect of CLA are carried out through that transcription factor (Khan and Vanden-Heuvel, 2003), leading to increased β -oxidation and energy expenditure (Martin *et al.* 2000). In the study of Royan *et al.* (2011b), the PPAR α gene expression in the liver tissue of broiler chickens fed CLA was lower than in tissue in birds fed fish oil.

CLA effects on the chicken immune system

The first reports showed that dietary CLA might prevent the growth depression induced by immune stimulation in mice and chickens (Cook *et al.* 1993). However, in the study by Long *et al.* (2011), the immunoregulatory actions of CLA of relevance to viral disease pathogenesis and immune responses were investigated. Their results indicated that dietary CLA enhanced immune function in chickens, particularly those of the IBDV-immunosuppressive status. Furthermore, at the molecular level, the immunoregulatory functions of CLA on chickens were attributable mainly to the antiinflammatory properties of CLA and were mediated, at least in part, by suppressing the IBDV-specific proinflammatory cytokine mRNA relative expression. Long *et al.* (2012) investigated the immunoregulatory actions of CLA and suggested that CLA alleviated the immunosuppression of T lymphocytes in broiler chickens exposed to cyclosporine A through increased peripheral blood T lymphocyte proliferation and interleukin-2 levels.

CLA effects on chicken performance

There is no consistency in previous reports concerning the effects of CLA on the chicken's weight gain. Thiel-Cooper *et al.* (2001) found a linear increase of daily weight gain associated with CLA supplementation. There are also reports on the moderate weight loss caused by the dietary CLA in chickens (Cook *et al.* 1993) and in mice (Miller *et al.* 1994) exposed to

catabolic stress caused by endotoxin injections. In the study of Suksombat *et al.* (2007), the daily feed intake was not different in 3 week old broiler chickens fed up to 1.5% dietary CLA-compared to the control group; nevertheless, a lower daily weight gain was observed in CLA fed chicks. They also reported a decrease in abdominal fat and in drumstick and in boneless drumstick percents, along with higher liver weight in chicks fed CLA containing diets. Similar results on liver weight were also reported by Leaflet (2004).

Royan *et al.* (2011a) reported that broilers fed a high dietary CLA dose (4.2%), were found to have lower weight gains, but the chicks fed diets containing 2.1% CLA showed higher body weight gain than those fed 4.2% CLA in finisher diets. The adverse effect of CLA on body weight gain was reported by Szymczyk *et al.* (2001) and Suksombat *et al.* (2007) with dietary CLA levels up to 1.5%. Similar results were reported by Buccioni *et al.* (2009) using 1% dietary CLA; however, in their research, the weight gains of treated birds fluctuated and were lower at the initial and the final phases, but higher during the mid phase of the experiment. In the study of Royan *et al.* (2011a), the 4.2% CLA diet reduced feed intake during the grower phase, but birds recovered it later in the finisher phase. On the other hand the 2.1% dietary CLA level did not affect feed intake. It seems that the ability of chickens to use CLA increases with age, so that Suksombat *et al.* (2007) found that the feed intake of broiler chickens was significantly decreased by dietary CLA over the starter period, while no effects of CLA on this variable were noted over the grower-finisher period. In spite of that, the feed intake depression in the whole experimental period was statistically significant.

In most previous reports, feed intake was unaffected by the incorporation of CLA into broiler chickens diets. These observations were recorded using different dietary CLA levels: 0.4% (Denli *et al.* 2004), 1% (Takahashi *et al.* 2003; Zhang *et al.* 2005; Buccioni *et al.* 2009), 1.5% (Szymczyk *et al.* 2001; Suksombat *et al.* 2007) 1.8% (Simon *et al.* 2000), 2% (Bolukbasi, 2006) 3% (Du and Ahn, 2003) and 4% (Sirri *et al.* 2003). In other reports, Du and Ahn (2002) showed that up to 1% dietary CLA had no effect on feed consumption, but 2% and 3% levels reduced feed consumption of broilers. Also in the study by Javadi *et al.* (2007), the 1% dietary CLA was enough to reduce broiler chicken feed intake. The only report of a positive effect of CLA on feed intake is that of Bolukbasi (2006), who reported a higher feed intake in broiler chickens fed diets containing 1% CLA compared with the control group, but they found no difference between dietary 2 and 3% CLA levels and the control diet. In the study of Royan *et al.* (2011a) the diets containing CLA unfavorably increased FCR. The variations in FCR clearly resulted from differences in body weight gain, which is consistent with the report of Suksombat *et al.* (2007). However, most reports fail to demonstrate the existence of an effect of the dietary CLA on the feed

conversion ratio of broiler chickens (Simon *et al.* 2000; Javadi *et al.* 2007; Zhang *et al.* 2005; Szymczyk *et al.* 2001; Du and Ahn, 2002).

Moreover, a lower feed conversion ratio (which means higher efficiency) was observed in diets supplemented by CLA (Sell *et al.* 2001; Szymczyk *et al.* 2001; Bolukbasi, 2006), suggesting that the pattern of CLA effects in broiler chickens performance is highly variable among studies. However, it seems that the growth rate was more susceptible to CLA unfavorable effects than other performance traits. Apparently, CLA inclusion in diets at levels above 10 g/kg decreased the broilers growth rate (Szymczyk *et al.* 2001; Badinga *et al.* 2003). Royan *et al.* (2011a) found that the negative effects of CLA on growth rate are somewhat modified by dietary CLA dose, age of birds and the fat composition of the experimental diets, so that the chicks fed diets containing 2.1% CLA in the finisher phases showed an acceptable body weight gain compared to the 4.2% CLA level. The combination of CLA + soybean oil resulted in higher weight gain than the combination of CLA + fish oil.

Adverse results have also been reported on the effects of CLA on layer hen's performance. Szymczyk and Pisulewski (2003) reported that the feed intake and mass of eggs produced by hens fed with CLA-enriched diets were lower than those recorded for the control group. Thus, since the mass of eggs was more affected by dietary CLA, the resulting average feed conversion ratio, expressed as kg of feed required per kg of eggs produced, was higher in hens fed the CLA-enriched diets than in those fed the control diet. Cho *et al.* (2013) investigated the effect of CLA feeding on growth performance and fatty acid profiles in thigh meat of broiler chicken using meta-analysis on a total of 9 studies. They concluded that CLA was not beneficial for improving growth performance, although it might be estimated that CLA is effective in modulating n-6/n-3 fatty acids ratio in thigh meat. However a comparison of the loss from suppressed growth performance and increased saturated fatty acids with the benefit from enhanced n-6/n-3 ratio should be investigated in further studies in order to propose the appropriate use of dietary CLA in the broiler industry.

Effects of CLA on chicken carcass traits

The only report on the positive effects of CLA on carcass yield is that of Buccioni *et al.* (2009) where broiler chickens were fed diets with 1% CLA. Royan *et al.* (2011a) observed some differences in carcass yield (after feathers and gut had been removed) between CLA containing diets and other treatments. The combination of soybean oil with CLA (2.1% CLA+3.5% soybean oil) prevented the carcass yield depression caused by the diets containing 4.2% CLA. The breast percentage changed in the same manner as described above.

In the majority of the previous studies, carcass traits were not affected by dietary CLA. Bolukbasi (2006) did not

found differences in the carcass yield and leg weight of chicks fed diets containing up to 3% CLA combined with sunflower oil, but they found an increase in the breast percentage in birds fed CLA. In the study of Szymczyk *et al.* (2001), the relative proportion of breast and leg muscles (% of carcass weight) responded differently to increasing levels of dietary CLA. The former variable was not affected by the treatment and the latter was significantly increased.

In other previous reports, the dietary inclusion of CLA at 0.4% (Denli *et al.* 2005), 1.5% (Suksombat *et al.* 2007) or 4% (Sirri *et al.* 2003) levels did not alter dressing percentage or thigh and breast yields. Buccioni *et al.* (2009) claimed that the increase in dressing percentage in CLA treated animals was related to a significant decrease in abdominal separable fat, and attributed such effect to the ability of CLA to reduce body fat accretion. In a comparable report, Suksombat *et al.* (2007) showed that the reduced abdominal fat pad in birds fed dietary CLA was not accompanied by an increase in carcass, breast or thigh percents. These controversies suggest that the observed CLA effect on carcass parameters of broiler chickens is not simply the result of abdominal fat pad alterations.

Effects of CLA on abdominal fat pad

There are some conflicting reports on the effects of CLA on the abdominal fat pad changes in chickens. Du and Ahn (2002) found that feeding a diet containing 0.5% CLA to broilers at 3 weeks of age, for a period of 3 weeks, resulted in an increase in abdominal fat content. Similar results have been reported by Denli *et al.* (2005) and Javadi *et al.* (2007). Nevertheless, there are also some conflicting reports concerning the effects of CLA on abdominal fat pad reduction in chickens (Simon *et al.* 2000; Szymczyk *et al.* 2001; Badinga *et al.* 2003). It was reported that CLA is effective in reducing body fat deposition only during the weight gain phase (at the earlier ages when a positive energy balance exists) (Atkinson, 1999; Kamphuis *et al.* 2003a; Kamphuis *et al.* 2003b; Larsen *et al.* 2006).

It seems that the CLA effect on fat accumulation can be modified by the associated of dietary fat supplements. Zanini *et al.* (2006) demonstrated a linear reduction in abdominal fat pad when canola oil was used with CLA, but not when CLA was combined with soybean oil. In another report, it was shown that CLA, in combination with coconut oil or soybean oil (SFA and n-6 PUFA rich, respectively), could decrease body fat mass in mice (Kennedy, 2007); still, mice fed CLA in combination with fish oil (n-3 PUFA rich) showed no effect on adiposity (Hargrave *et al.* 2005). According to the review of Park and Pariza (2007) the effects of CLA on body composition can be attributed to: the rise in energetic expenditure through the increased synthesis of uncoupling proteins; the decrease of body fat deposits

through a reduction in the number and size of adipocytes due to the inhibition of lipoprotein lipase enzyme, the stimulation of the apoptosis process of pre-adipocytes and the increase in lipolysis and β -oxidation in muscle tissue as indicated by higher carnitine acyltransferase I and II.

Effects of CLA on the broiler's liver

An increase in liver weight of broiler chickens fed diets containing CLA has been reported (Du and Ahn, 2003; Leaflet, 2004; Suksombat *et al.* 2007; Royan *et al.* 2011a). In the report of Buccioni *et al.* (2009), the change in liver weight were dose-dependent, so that the 0.5% dietary CLA significantly increased liver weight but the 1% and 1.5% doses did not result in any difference compared with the control diet. Zanini *et al.* (2006) showed a reduction in the relative liver weight in female broiler chickens supplied with CLA and canola oil (n-3 rich). They found an interaction between oil source and CLA such that supplementation of CLA produced lower relative liver weight in birds fed canola oil compared to that of birds receiving CLA + soybean oil in their diet.

Based on animal studies, one main concern about CLA use, identified so far, is fatty liver (Pariza, 2004). In the study of Royan *et al.* (2013) the CLA containing diets increased liver fat accumulation and the diet with 4.2% CLA had a larger effect than diets containing 2.1% CLA. Fatty liver may be the result of CLA's pronounced effects on body fat mobilization, as well as increased fatty acid synthesis in the liver (Clement *et al.* 2002; Tsuboyama-Kasaoka *et al.* 2000; Yanagita *et al.* 2005). Zanini *et al.* (2006) reported an interaction between n-3 and n-6 fatty acids in relation to dietary CLA. In their research, adding CLA to the diets containing soybean oil resulted in an increase in the total lipid content of the liver, but the opposite was observed for the combination of canola oil and CLA. Nevertheless without CLA supplementation, liver fat content was higher for birds fed canola oil compared with those fed a diet containing soybean oil.

Effect of CLA on the chicken's serum parameters

The effect of dietary CLA on increasing serum TG concentration was reported by Du and Ahn (2003). However other studies report a decrease in serum or plasma TG following CLA administration (Bolukbasi, 2006). The reason for the higher serum TG concentration in birds fed dietary CLA is not clear, but it could be attributed to changes in the activities of enzymes involved in hepatic lipid metabolism. In birds, lipid synthesis occurs mainly in the liver. This effect may be a consequence of the inhibitory role of CLA on lipoprotein lipase together with stimulation of lipolysis in adipose tissue (Park *et al.* 1997). Consequently, reduced fat deposits and higher lipolysis in adipocytes could be the

reason for the elevated serum TG levels, as observed in broiler chickens. In the study of Du and Ahn (2003) dietary CLA caused a significant increase in liver fatty acid synthase (FA synthase) activity and an increase (even though not significant) in acetyl-CoA carboxylase activity. FA synthase and acetyl-CoA carboxylase are the main enzymes regulating fatty acid synthesis. The higher FA synthase activity could be explained in part by the increased plasma TG levels. In the mammary glands of sows, dietary CLA caused a reduction in acetyl-CoA carboxylase and FA synthase (Piperova *et al.* 2000), but in cultured adipose cells, FA synthase gene expression was not decreased by dietary CLA (Choi *et al.* 2000). These results show that dietary CLA decreases lipogenesis in mammary glands and adipose tissues but not in liver. This could be the explanation for the ineffectiveness of CLA in decreasing fat deposition in birds (Du and Ahn 2002), because liver is the main site of lipogenesis. Fatty acid synthesis in mice, rats and pigs takes place mainly in adipose tissues, and the lipogenesis inhibitory effect of CLA in adipose tissue could considerably decrease fat accumulation in these species. The coordinated change of total cholesterol and LDL in birds fed CLA has been reported (Bhattacharya *et al.* 2006; Baddini Feitoza *et al.* 2009; Stangl *et al.* 1999), along with an increase in serum HDL level in broiler chickens fed CLA (Bolukbasi, 2006; Denli *et al.* 2005; Du and Ahn, 2003).

CONCLUSION

This review showed that CLA has the potential to modify metabolism and metabolic rate in broilers, but CLA feeding can also reduce productivity and increase SFA levels in chicken meat. There is no consistency in previous reports concerning the effects of different dietary dosages of CLA on chicken's performance, but the levels above 1% are usually more risky. Therefore, CLA can be useful as an additive to produce meat rich in n-3 PUFA and to lower the n-6/n-3 ratio. A balance between improved meat quality and decreased growth performance is needed and the customer's preference should also be considered.

REFERENCES

- Adolf R., Duval S. and Emeken E. (2000). Biosynthesis of conjugated linoleic acid in humans. *Lipids*. **35**, 131-135.
- Atkinson R.L. (1999). Conjugated linoleic acid for altering body composition and treating obesity. Pp. 348-353 in *Advances in Conjugated Linoleic Acid Research*. M.P. Yurawecz, M.M. Mossoba, J.K.G. Kramer, M.W. Pariza and G.J. Nelson, Eds. AOCS Press, Champaign, IL, USA.
- Aydin R., Pariza M. and Cook M. (2001). Olive oil prevents the adverse effects of dietary conjugated linoleic acid on chick hatchability and egg quality. *J. Nutr.* **131**, 800-806.

- Azain M.J., Hausman D.B., Sisk M.B., Flatt W.P. and Jewell D.E. (2000). Dietary conjugated linoleic acid reduces rat adipose tissue cell size rather than cell number. *J. Nutr.* **130**, 1548-1554.
- Baddini F.A., Pereira N.F., Da Costa F. and Ribeiro B.G. (2009). Conjugated linoleic acid (CLA): effect modulation of body composition and lipid profile. *Nutr. Hospital.* **24**, 422-428.
- Badinga L., Selberg K.T., Dinges A.C., Comer C.W. and Miles R.D. (2003). Dietary conjugated linoleic acid alters hepatic lipid content and fatty acid composition in broiler chickens. *Poult. Sci.* **82**, 111-116.
- Banni S. (2002). Conjugated linoleic acid metabolism. *Curr. Opin. Lipidol.* **13**, 261-266.
- Banni S. and Martin J.C. (1998). Conjugated linoleic acid and metabolites. Pp. 261-302 in *Trans Fatty Acids in Human Nutrition*. W.W. Christie and J. L. Se' be' dio, Eds. The Oily Press, Dundee, Scotland.
- Bassaganya-Riera J. and Hontecillas R. (2006). CLA and n-3 PUFA differentially modulate clinical activity and colonic PPAR-responsive gene expression in a pig model of experimental IBD. *Clin. Nutr.* **25**, 454-465.
- Bassaganya-Riera J., Pogranichniy R.M., Jobgen S.C., Halbur P.G., Yoon K.J., O'Shea M. and Hontecillas R. (2003). Conjugated linoleic acid ameliorates viral infectivity in a pig model of virally induced immunosuppression. *J. Nutr.* **133**, 3204-3214.
- Berge G.M., Ruyter B. and Asgard T. (2004). Conjugated linoleic acid in diets for juvenile Atlantic salmon (*Salmosalar*); effects on fish performance, proximate composition, fatty acid and mineral content. *Aquaculture.* **237**, 365-380.
- Bhattacharya A., Banu J., Rahman M., Causey J. and Fernandes G. (2006). Biological effects of conjugated linoleic acids in health and disease. *J. Nutr. Biochem.* **17**, 789-810.
- Bolkubasi S.C. (2006). Effect of dietary conjugated linoleic acid (CLA) on broiler performance, serum lipoprotein content, muscle fatty acid composition and meat quality during refrigerated storage. *Br. Poult. Sci.* **47**, 470-476.
- Bretillon L., Chardigny J.M., Gregoire O., Berdeaux O. and Sebedio J.L. (1999). Effects of conjugated linoleic acid isomers on the hepatic microsomal desaturation activities *in vitro*. *Lipids.* **34**, 956-969.
- Brodie A.E., Manning V.A., Ferguson K.R., Jewell D.E. and Hu C.Y. (1999). Conjugated linoleic acid inhibits differentiation of pre- and post confluent 3T3-L1 preadipocytes but inhibits cell proliferation only in pre-confluent cells. *J. Nutr.* **129**, 602-606.
- Brown J.M., Boysen M.S., Jensen S.S., Morrison R.F., Storkson J., Lea-Currie R., Pariza M., Mandrup S. and McIntosh M.K. (2003). Isomer-specific regulation of metabolism and PPARs signaling by CLA in human preadipocytes. *J. Lipid. Res.* **44**, 1287-1300.
- Brown J.M., Halvorsen Y.D., Lea-Currie Y.R., Geigerman C. and McIntosh M. (2001). Trans-10, cis-12, but not cis-9, trans-11, conjugated linoleic acid attenuates lipogenesis in primary cultures of stromal vascular cells from human adipose tissue. *J. Nutr.* **131**, 2316-2321.
- Boutheadour J.C., Evans P.C., Grippo D., Tiffon B., Blouguit M.E., Roseau S., Lutton C., Tombe D. and Martin J.C. (2002). A CLA mixture prevents body triglyceride accumulation without affecting energy expenditure in Syrian hamsters. *J. Nutr.* **132**, 2682-2689.
- Buccioni A., Antongiovanni M., Mele M., Gualtieri M., Minieri S. and Rapaccini S. (2009). Effect of oleic and conjugated linoleic acid in the diet of broiler chickens on the live growth performances, carcass traits and meat fatty acid profile. *Italian J. Anim. Sci.* **8**, 603-614.
- Changhua L., Jindong Y., Defa L., Lidan Z., Shiyun Q. and Jianjun X. (2005). Conjugated linoleic acid attenuates the production and gene expression of proinflammatory cytokines in weaned pigs challenged with lipopolysaccharide. *J. Nutr.* **135**, 239-244.
- Chin S.F., Liu W., Storkson J.M., Ha Y.L. and Pariza M.W. (1992). Dietary sources of conjugated dienoic isomers of linoleic acid, a newly recognized class of anticarcinogens. *J. Food Comp. Anal.* **5**, 185-197.
- Chin S.F., Storkson J.M., Albright K.J., Cook M.E. and Pariza M.W. (1994). Conjugated linoleic acid is a growth factor for rats as shown by enhanced weight gain and improved feed efficiency. *J. Nutr.* **124**, 2344-2349.
- Cho S., Ryua C., Yang J., Mbiriri D.T., Choi C.W., Chae J.I., Kim Y.H., Shim K.S., Kim Y.J. and Choi N.J. (2013). Effect of conjugated linoleic acid feeding on the growth performance and meat fatty acid profiles in broiler: meta-analysis. *Asian-Australas J. Anim. Sci.* **26**, 995-1002.
- Choi J.S., Jung M.H., Park H.S. and Song J. (2004). Effect of conjugated linoleic acid isomers on insulin resistance and mRNA levels of genes regulating energy metabolism in high-fat-fed rats. *Nutrition.* **20**, 1008-1017.
- Choi Y., Kim Y.C., Han Y.B., Park Y., Pariza M.W. and Ntambi J.M. (2000). The trans-10, cis-12 isomer of conjugated linoleic acid down regulates stearyl-CoA desaturase gene expression in 3T3-L1 adipocytes. *J. Nutr.* **130**, 1920-1924.
- Choi Y., Park Y., Storkson J.M., Pariza M.W. and Ntambi J.M. (2002). Inhibition of stearyl-CoA desaturase activity by the cis-9, trans-11 isomer and the trans-10, cis-12 isomer of conjugated linoleic acid in MDA-MB-231 and MCF-7 human breast cancer cells. *Biochem. Biophys. Res. Commun.* **294**, 785-790.
- Clement L., Poirier H., Niot I., Bocher V., Guerre-Millo M., Krief S., Staels B. and Besnard P. (2002). Dietary trans-10, cis-12 conjugated linoleic acid induces hyperinsulinemia and fatty liver in the mouse. *J. Lipid. Res.* **43**, 1400-1409.
- Cohen C., Perrault G., Voltz C., Steinberg R. and Soubrié P. (2002). SR141716, a central cannabinoid (CB1) receptor antagonist, blocks the motivational and dopamine-releasing effects of nicotine in rats. *Behav. Pharmacol.* **13**, 451-463.
- Cook M.E., Jerome D. and Pariza M.W. (2000). Method for selectively altering body fat level, feed efficiency, or weight gain. *US Patent.* **6**, 378.
- Cook M.E., Miller C.C., Park Y. and Pariza M.W. (1993). Immune modulation by altered nutrient metabolism: nutritional control of immune-induced growth depression. *Poult. Sci.* **72**, 1301-1305.
- Corl B.A., Baumgard L.H., Dwyer D.A., Griinari J.M., Phillips B.S. and Bauman D.E. (2001). The role of delta (9)-desaturase in the production of cis-9, trans-11 CLA. *J. Nutr. Biochem.* **12**, 622-630.
- Degrace P., Demizieux L., Gresti J., Chardigny J.M., Sébédio J.L. and Clouet P. (2004). Hepatic steatosis is not due to impaired fatty acid oxidation capacities in C57BL/6J mice fed the conjugates trans-10, cis-12-isomer of linoleic acid. *J. Nutr.* **134**, 861-867.
- Denli M., Okan F. and Doran F. (2004). Effect of conjugated linoleic acid (CLA) on the performance and serum variables of broiler chickens intoxicated with aflatoxin B₁. *South African J. Anim. Sci.* **34**, 97-103.

- Denli M., Okan F., Doran F. and Inal T.C. (2005). Effect of dietary conjugated linoleic acid (CLA) on carcass quality, serum lipid variables and histopathological changes of broiler chickens infected with aflatoxin B₁. *South African J. Anim. Sci.* **35**, 109-116.
- DeLany J.P., Blohm F., Truett A.A., Scimeca J.A. and West D.B. (1999). Conjugated linoleic acid rapidly reduces body fat content in mice without affecting energy intake. *Am. J. Physiol.* **276**, 1172-1179.
- Du M. and Ahn D.U. (2002). Effect of dietary conjugated linoleic acid on the growth rate of live birds and on the abdominal fat content and quality of broiler meat. *Poult. Sci.* **81**, 428-433.
- Du M. and Ahn D.U. (2003). Dietary CLA affects lipid metabolism in broiler chicks. *Lipids*. **38**, 505-511.
- Engelke C.F., Siebert B.D., Gregg K., Wright A.D.G. and Vercoe P.E. (2004). Kangaroo adipose tissue has higher concentrations of cis 9, trans 11-conjugated linoleic acid than lamb adipose tissue. *J. Anim. Feed. Sci.* **13**, 689-692.
- Evans M., Brown J. and McIntosh M. (2002). Isomer-specific effects of conjugated linoleic acid (CLA) on adiposity and lipid metabolism. *J. Nutr. Biochem.* **13**, 508-516.
- Evans M., Geigerman C., Cook J., Curtis L., Kuebler B. and McIntosh M. (2000). Conjugated linoleic acid suppresses triglyceride accumulation and induces apoptosis in 3T3-L1 preadipocytes. *Lipids*. **35**, 899-910.
- Fogerty A.C., Ford G.L. and Svoronos D. (1988). Octadeca-9, 11-dienoic acid in foodstuffs and in the lipids of human blood and breast milk. *Nutr. Reports. Int.* **38**, 937-944.
- Fritsche J. and Steinhart C. (1998). Amounts of conjugated linoleic acid (CLA) in German foods and evaluation of daily intake. *Zeitschrift Lebensm. Unters. Forsch.* **206**, 77-82.
- Gregoire F.M., Smas C.M. and Sul H.S. (1998). Understanding adipocyte differentiation. *Physiol. Rev.* **78**, 783-809.
- Griinari J.M. and Bauman D.E. (1999). Biosynthesis of conjugated linoleic acid and its incorporation into meat and milk in ruminants. Pp. 180-200 in *Advances in Conjugated Linoleic Acid Research*. M.P. Yurawecz, M.M. Mossoba, J.K.G. Kramer, M.W. Pariza and G.J. Nelson, Eds. AOCS Press, Champaign, IL.
- Griinari J.M., Corl B.A., Lacy S.H., Chouinard P.Y., Nurmela K.V. and Bauman D.E. (2000). Conjugated linoleic acid is synthesized endogenously in lactating dairy cows by delta (9)-desaturase. *J. Nutr.* **130**, 2285-2291.
- Ha Y.L., Grimm N.K. and Pariza M.W. (1987). Anticarcinogens from fried ground beef: heat-altered derivatives of linoleic acid. *Carcinogenesis*. **8**, 1881-1887.
- Hargrave K.M., Azain M.J. and Miner J.L. (2005). Dietary coconut oil increases conjugated linoleic acid-induced body fat loss in mice independent of essential fatty acid deficiency. *Biochem. Biophys. Acta.* **1737**, 52-60.
- Herzallah S. (2013). Enrichment of conjugated linoleic acid (CLA) in hen eggs and broiler chickens meat by lactic acid bacteria. *Br. Poult. Sci.* **54**, 747-752.
- Hotamisligil G.S. and Spiegelman B.M. (1994). Tumor necrosis factor alpha: a key component of the obesity-diabetes link. *Diabetes*. **43**, 1271-1278.
- Houseknecht K.L., VandenHeuvel J.P., Moya-Camarena S.Y., Portocarrero C.P., Peck L.W., Nickel K.P. and Belury M.A. (1998). Dietary conjugated linoleic acid normalizes impaired glucose tolerance in the Zucker diabetic fatty fa/fa rat. *Biochem. Biophys. Res. Commun.* **244**, 678-682.
- Hur S.J., Yang H.S., Park G.B. and Joo S.T. (2007). Effects of dietary glycine betaine on pork quality in different muscle types. *Asian-Australas J. Anim. Sci.* **20**, 1754-1760.
- Inoue N., Nagao K., Hirata J., Wang Y.M. and Yanagita T. (2004). Conjugated linoleic acid prevents the development of essential hypertension in spontaneously hypertensive rats. *Biochem. Biophys. Res. Commun.* **323**, 679-684.
- Ip C., Dong Y., Ip M.M., Banni S., Carta G., Angioni E., Murru E., Spada S., Melis M.P. and Saebo A. (2002). Conjugated linoleic acid isomers and mammary cancer prevention. *Nutr. Cancer*. **43**, 52-58.
- Javadi M., Math J.H., Everts G.H., Hovenier R., Javadi S., Kappert H. and Beynen A.C. (2007). Effect of dietary conjugated linoleic acid on body composition and energy balance in broiler chickens. *Br. J. Nutr.* **98**, 1152-1158.
- Joo S.T., Lee J.I., Ha Y.L. and Park G.B. (2002). Effects of dietary conjugated linoleic acid on fatty acid composition, lipid oxidation, color, and water-holding capacity of pork loin. *J. Anim. Sci.* **80**, 108-112.
- Kamphuis M.M., Lejeune M.P., Saris W.H. and Westerterp-Plantenga M.S. (2003a). Effect of conjugated linoleic acid supplementation after weight loss on appetite and food intake in overweight subjects. *European J. Clin. Nutr.* **57**, 1268-1274.
- Kamphuis M.M., Lejeune M.P., Saris W.H. and Westerterp-Plantenga M.S. (2003b). The effect of conjugated linoleic acid supplementation after weight loss on body weight regain, body composition, and resting metabolic rate in overweight subjects. *Int. J. Obesity. Rel. Metab. Disord.* **27**, 840-847.
- Kawahara S., Takenoyama S., Takuma K., Muguruma M. and Yamauchi K. (2009). Effects of dietary supplementation with conjugated linoleic acid on fatty acid composition and lipid oxidation in chicken breast meat. *Anim. Sci. J.* **80(4)**, 468-474.
- Kelley N.S., Hubbard N.E. and Erickson K.L. (2007). Conjugated linoleic acid isomers and cancer. *J. Nutr.* **137**, 2599-2607.
- Kennedy S.R. (2007). Bioactive fatty acids as dietary supplements for farmed fish: effects on growth performance, lipid metabolism, gene expression and immune parameters. Ph D. Thesis. University of Stirling, Scotland.
- Kepler C.R., Hirons K.P., McNeill J.J. and Tove S.B. (1966). Intermediates and products of the biohydrogenation of linoleic acid by *Butyrivibrio fibrisolvens*. *J. Biol. Chem.* **241**, 1350-1354.
- Khan S.A. and Vanden Heuvel J.P. (2003). Role of nuclear receptors in the regulation of gene expression by dietary fatty acids: a review. *J. Nutr. Biochem.* **14**, 554-567.
- Khanal R.C. and Dhiman T.R. (2004). Biosynthesis of conjugated linoleic acid (CLA): a review. *Pakistan J. Nutr.* **3**, 72-81.
- Kim H.Y. (2007). Novel metabolism of docosahexaenoic acid in neural cells. *J. Biol. Chem.* **282**, 18661-18665.
- Knekt P., Jarvinen R., Seppanen R., Pukkala E. and Aromaa A. (1996). Intake of dairy products and the risk of breast cancer. *Br. J. Cancer.* **73**, 687-691.
- Knight T.W., Knowles S., Death A.F., West J., Agnew M., Morris C.A. and Purchas R.W. (2003). Factors affecting the variation in fatty acid concentrations in lean beef from grass-fed cattle in New Zealand and the implications for human health. *New Zealand J.*

- Agric. Res.* **46**, 83-95.
- Kramer J.K.G., Sehat N., Dugan M.E.R., Mossoba M.M., Yurawecz M.P. and Roach J.A.G. (1998). Distributions of conjugated linoleic acid (CLA) isomers in tissue lipid classes of pigs fed a commercial CLA mixture determined by gas chromatography and silver ion-high-performance liquid chromatography. *Lipids*. **33**, 549-558.
- Kritchevsky D. (2000). Antimutagenic and some other effects of conjugated linoleic acid. *Br. J. Nutr.* **83**, 459-465.
- Larsen T.M., Toubro S., Gudmundsen O. and Astrup A. (2006). Conjugated linoleic acid supplementation for 1 y does not prevent weight or body fat regain. *Am. J. Clin. Nutr.* **83**, 606-612.
- Larsson S.C., Bergkvist L. and Wolk A. (2005). High-fat dairy food and conjugated linoleic acid intakes in relation to colorectal cancer incidence in the Swedish Mammography Cohort. *Am. J. Clin. Nutr.* **82**, 894-900.
- Leaflet A.S. (2004). Dietary Conjugated Linoleic Acid (CLA) Effects Lipid Metabolism in Broiler Chicks R1934: Iowa State University Animal Industry Report.
- Lee K.N., Pariza M.W. and Ntambi J.M. (1998). Conjugated linoleic acid decreases hepatic stearoyl-CoA desaturase mRNA expression. *Biochem. Biophys. Res. Com.* **248**, 817-821.
- Lee K.W., Lee H.J., Cho H.Y. and Kim Y.J. (2005). Role of the conjugated linoleic acid in the prevention of cancer. *Crit. Rev. Food Sci. Nutr.* **45**, 135-144.
- Lin Y., Kreeft A., Schuurbiens J.A.E. and Draijer R. (2001). Different effects of conjugated linoleic acid isomers on lipoprotein lipase activity in 3T3-L1 adipocytes. *J. Nutr. Biochem.* **12**, 183-189.
- Long F.Y., Guo Y.M., Wang Z., Liu D., Zhang B.K. and Yang X. (2011). Conjugated linoleic acids alleviate infectious bursal disease virus-induced immunosuppression in broiler chickens. *Poult. Sci.* **90**, 1926-1933.
- Long F.Y., Yang X., Guo Y.M., Wang Z., Yuan J.M., Zhang B.K. and Liu D. (2012). Conjugated linoleic acids alleviate the immunosuppression of peripheral blood T lymphocytes in broiler chickens exposed to cyclosporin A. *Poult. Sci.* **91**, 2431-2437.
- Martin J.C., Grégoire S., Siess M.H., Genty M., Chardigny J.M., Berdeaux O., Juaneda P. and Sébédio J.L. (2000). Effects of conjugated linoleic acid isomers on lipid-metabolizing enzymes in male rats. *Lipids*. **35**, 91-98.
- Masso-Welch P.A., Zangani D., Ip C., Vaughan M.M., Shoemaker S.F., McGee S.O. and Ip M.M. (2004). Isomers of conjugated linoleic acid differ in their effects on angiogenesis and survival of mouse mammary adipose vasculature. *J. Nutr.* **134**, 299-307.
- Masso-Welch P.A., Zangani D., Ip C., Vaughan M.M., Shoemaker S., Ramirez R.A. and Ip M.M. (2002). Inhibition of angiogenesis by the cancer chemopreventive agent conjugated linoleic acid. *Cancer Res.* **62**, 4383-4389.
- McLeod R.S., LeBlanc A.M., Langille M.A., Mitchell P.L. and Currie D.L. (2004). Conjugated linoleic acids, atherosclerosis and hepatic very-low-density lipoprotein metabolism. *Am. J. Clin. Nutr.* **79**, 1169-1174.
- Mersmann H.J. (2002). Mechanisms for conjugated linoleic acid-mediated reduction in fat deposition. *J. Anim. Sci.* **80**, 126-134.
- Miller C.C., Park Y., Pariza M.W. and Cook M.E. (1994). Feeding conjugated linoleic acid to animals partially overcomes catabolic response due to endotoxin injection. *Biochem. Biophys. Res. Commun.* **198**, 1107-1112.
- Mosley E.E., McGuire M.K., Williams J.E. and McGuire M.A. (2006). Cis-9, trans-11 conjugated linoleic acid is synthesized from vaccenic acid in lactating women. *J. Nutr.* **136**, 2297-2301.
- Moya-Camarena S.Y., Heuvel J.P.V. and Blanchard S.G. (1999). Conjugated linoleic acid is a potent naturally occurring ligand and activator of PPAR α . *J. Lipid. Res.* **40**, 1426-1433.
- Muller H.L., Kirchgessner M., Roth F.X. and Stangl G.I. (2000). Effect of conjugated linoleic acid on energy metabolism in growing-finishing pigs. *J. Anim. Physiol. Anim. Nutr.* **83**, 85-94.
- Muller H.L., Stangl G.I. and Kirchgessner M. (1999). Energy balance of conjugated linoleic acid-treated pigs. *J. Anim. Physiol. Anim. Nutr.* **81**, 150-156.
- Nagao K., Inoue N., Wang Y.M., Hirata J., Shimada Y., Nagao T., Matsui T. and Yanagita T. (2003). The 10 trans, 12 cis isomer of conjugated linoleic acid suppresses the development of hypertension in Otsuka Long-Evans Tokushima fatty rats. *Biochem. Biophys. Res. Com.* **306**, 134-138.
- Nagao K., Inoue N., Wang Y.M., Shirouchi B. and Yanagita T. (2005). Dietary conjugated linoleic acid alleviates nonalcoholic fatty liver disease in Zucker (fa/fa) rats. *J. Nutr.* **135**, 9-13.
- Nagao K., Inoue N., Wang Y.M. and Yanagita T. (2003). Conjugated linoleic acid enhances plasma adiponectin level and alleviates hyperinsulinemia and hypertension in Zucker diabetic fatty (fa/fa) rats. *Biochem. Biophys. Res. Commun.* **310**, 562-566.
- Ohnuki K., Haramizu S., Ishihara K. and Fushiki T. (2001). Increased energy metabolism and suppressed body fat accumulation in mice by a low concentration of conjugated linoleic acid. *Biosci. Biotechnol. Biochem.* **65**, 2200-2204.
- Pariza M.W., Park Y. and Cook M.E. (2000). Mechanisms of action of conjugated linoleic acid, evidence and speculation. *Proc. Soc. Exp. Biol. Med.* **223**, 8-13.
- Pariza M.W., Park Y. and Cook M.E. (2001). The biologically active isomers of conjugated linoleic acid. *Prog. Lipid. Res.* **40**, 283-298.
- Pariza M.W. (2004). Perspective on the safety and effectiveness of conjugated linoleic acid. *Am. J. Clin. Nutr.* **79**, 1132-1136.
- Pariza M.W. and Hargraves W.A. (1985). A beef-derived mutagenesis modulator inhibits initiation of mouse epidermal tumors by 7, 12-dimethylbenz [a] anthracene. *Carcinogenesis*. **6**, 591-593.
- Pariza M.W., Ashoor S.H., Chu F.S. and Lund D.B. (1979). Effects of temperature and time on mutagen formation in pan-fried hamburger. *Cancer. Letters.* **7**, 63-69.
- Park Y., Albright K.J., Liu W., Storkson J.M., Cook M.E. and Pariza M.W. (1997). Effect of conjugated linoleic acid on body composition of mice. *Lipids*. **32**, 853-858.
- Park Y., Albright K.J., Liu W., Storkson J.M., Lin M.E., Cook M.E. and Pariza M.W. (1999a). Changes in body composition in mice during feeding and withdrawal of conjugated linoleic acid. *Lipids*. **34**, 243-248.
- Park Y., Storkson J.M., Albright K.J., Liu W. and Pariza M.W. (1999b). Evidence that the trans-10, cis-12 isomer of conjugated linoleic acid induces body composition changes in mice. *Lipids*. **34**, 235-241.
- Park Y., Albright K.J., Storkson J.M., Liu W. and Pariza M.W. (2007). Conjugated linoleic acid (CLA) prevents body fat accumulation and weight gain in an animal model. *J. Food. Sci.* **72**, 612-617.
- Park Y. and Pariza M.W. (2007). Mechanisms of body fat modulation by conjugated linoleic acid (CLA). *Food Res. Int.* **40**, 311-323.

- Park Y., Storkson J.M., Liu W., Albright K.J., Cook M.E. and Pariza M.W. (2004). Structure-activity relationship of conjugated linoleic acid and its cognates in inhibiting heparin-releasable lipoprotein lipase and glycerol release from fully differentiated 3T3-L1 adipocytes. *J. Nutr. Biochem.* **15**, 561-568.
- Peters J.M., Park Y., Gonzalez F.J. and Pariza M.W. (2001). Influence of conjugated linoleic acid on body composition and target gene expression in peroxisome-proliferator-activated receptor α -Null Mice. *Biochem. Biophys. Acta.* **1533**, 233-241.
- Peterson D.G., Matitashvili E.A. and Bauman D.E. (2003). Diet-induced milk fat depression in dairy cows results in increased trans-10, cis-12 CLA in milk fat and coordinate suppression of mRNA abundance for mammary enzymes involved in milk fat synthesis. *J. Nutr.* **133**, 3098-3102.
- Piperova L.S., Teter B.B., Bruckental I., Sampugna J., Mills S.E., Yurawecz M.P., Fritsche J., Ku K. and Erdman R.A. (2000). Mammary lipogenic enzyme activity, trans fatty acids and conjugated linoleic acids are altered in lactating dairy cows fed a milk fat-depressing diet. *J. Nutr.* **130**, 2568-2574.
- Rahman S.M., Wang Y., Yotsumoto H., Cha J., Han S., Inoue S. and Yanagita T. (2001). Effects of conjugated linoleic acid on serum leptin concentration, body-fat accumulation and beta-oxidation of fatty acid in OLETF rats. *Nutrition.* **17**, 385-390.
- Royan M., Goh Y.M., Othman F., Sazili A.Q. and Navidshad B. (2011a). Effects of dietary conjugated linoleic acid (CLA), n-3 and n-6 fatty acids on performance and carcass traits of broiler chickens. *African J. Biotechnol.* **75**, 17379-17384.
- Royan M., Goh Y.M., Othman F., Sazili A.Q. and Navidshad B. (2011b). Effects of conjugated linoleic acid, fish oil and soybean oil on PPARs (α and γ) mRNA expression in broiler chickens and their relation to body fat deposits. *Int. J. Mol. Sci.* **12**, 8581-8595.
- Royan M., Meng G.Y., Othman F., Sazili A.Q. and Hanachi P. (2013). Effects of dietary combination of conjugated linoleic acid with fish oil or soybean oil on fatty acid composition of broiler meat. *Arch. Geflügelk.* **77**, 189-198.
- Santora J.E., Palmquist D.L. and Roehrig K.L. (2000). Trans-vaccenic acid is desaturated to conjugated linoleic acid in mice. *J. Nutr.* **130**, 208-215.
- Satory D.L. and Smith S.B. (1999). Conjugated acid inhibits proliferation but stimulates lipid filling of murine 3T3-L1 preadipocytes. *J. Nutr.* **129**, 92-97.
- Sell J.L., Jin S. and Jeffrey M. (2001). Metabolizable energy value of conjugated linoleic acid for broiler chicks and laying hens. *Poult. Sci.* **80**, 209-214.
- Shin D., Narciso-Gaytán C., Park J.H., Smith S.B., Sánchez-Plata M.X. and Ruiz-Feria C.A. (2011). Dietary combination effects of conjugated linoleic acid and flaxseed or fish oil on the concentration of linoleic and arachidonic acid in poultry meat. *Poult. Sci.* **90**, 1340-1347.
- Simon O., Manner K., Schafer K., Sagredos A. and Eder K. (2000). Effects of conjugated linoleic acids on protein to fat proportions, fatty acids, and plasma lipids in broilers. *European J. Lipid. Sci. Technol.* **102**, 402-410.
- Simon E., Marcarulla M.T., Fernandez-Quintela. A., Rodriguez V.M. and Portillo M.P. (2005). Body fat-lowering effect of conjugated linoleic acid is not due to increases lipolysis. *J. Physiol. Biochem.* **61**, 363-369.
- Sirri F., Minelli G., Iaffaldano N., Tallarico N. and Franchini A. (2003). Oxidative stability and quality traits of n-3 PUFA enriched chicken meats. *Italian J. Anim. Sci.* **2**, 450-452.
- Song H.J., Sneddon A.A., Barker P.A., Bestwick C., Choe S.N., McClinton S., Grant I., Rotondo D., Heys S.D. and Wahle K.W. (2004). Conjugated linoleic acid inhibits proliferation and modulates protein kinase C isoforms in human prostate cancer cells. *Nutr. Cancer.* **49**, 100-108.
- Stangl G.I., Muller H. and Kirchgessner M. (1999). Conjugated linoleic acid effects on circulating hormones, metabolites and lipoproteins, and its proportion in fasting serum and erythrocyte membranes of swine. *European J. Nutr.* **38**, 271-277.
- Storkson J.M., Park Y., Cook M.E. and Pariza M.W. (2005). Effects of trans-10, cis-12 conjugated linoleic acid (CLA) and cognates on apolipoprotein B secretion in HepG2 cells. *Nutr. Res.* **25**, 387-399.
- Suksombat W., Boonmee T. and Lounglawan P. (2007). Effects of various levels of conjugated linoleic acid supplementation on fatty acid content and carcass composition of broilers. *Poult. Sci.* **86**, 318-324.
- Szymczyk B. and Pisulewski P.M. (2003). Effects of dietary conjugated linoleic acid on fatty acid composition and cholesterol content of hen egg yolks. *Br. J. Nutr.* **90**, 93-99.
- Szymczyk B., Pisulewski P.M., Szczurek W. and Hanczakowski P. (2001). Effects of conjugated linoleic acid on growth performance, feed conversion efficiency and subsequent carcass quality in broiler chickens. *Br. J. Nutr.* **85**, 465-473.
- Takahashi K., Kawamata K., Akiba Y., Iwata T. and Kasai M. (2003). Effect of a mixture of conjugated linoleic acid isomers on growth performance and antibody production in broiler chicks. *Br. J. Nutr.* **89**, 691-694.
- Terpstra A.H., Beynen A.C., Everts H., Kocsis S., Katan M.B. and Zock P.L. (2002). The decrease in body fat in mice fed conjugated linoleic acid is due to increases in energy expenditure and energy loss in excreta. *J. Nutr.* **132**, 940-945.
- Thiel-Cooper R.L., Parrish F.C., Sparks J.C., Wiegand B.R. and Ewan R.C. (2001). Conjugated linoleic acid changes swine performance and carcass composition. *J. Anim. Sci.* **79**, 1821-182.
- Tontonoz P., Hu E., Graves R.A., Budavari A.I. and Spiegelman B.M. (1994). mPPAR- γ 2, tissue specific regulator of an adipocyte enhancer. *Genes. Dev.* **8**, 1224-1234.
- Tsuboyama-Kasaoka N., Takahashi M., Tanemura K., Kim H.J., Tange T., Okuyama H., Kasai M., Ikemoto S. and Ezaki O. (2000). Conjugated linoleic acid supplementation reduces adipose tissue by apoptosis and develops lipodystrophy in mice. *Diabetes.* **49**, 1534-1542.
- Turpeinen A.M., Mutanen M., Aro A., Salminen I., Basu S., Palmquist D.L. and Grünari J.M. (2002). Bioconversion of vaccenic acid to conjugated linoleic acid in humans. *Am. J. Clin. Nutr.* **76**, 504-510.
- Valeille K., Gripois D., Blouquit M.F., Souidi M., Riottot M., Bouthegourd J.C., Serougne C. and Martin J.C. (2004). Lipid atherogenic risk markers can be more favourably influenced by the cis-9, trans-11-octadecadienoate isomer than a conjugated linoleic acid mixture or fish oil in hamsters. *Br. J. Nutr.* **91**, 191-199.
- Wahle K.W.J., Heys S.D. and Rotondo D. (2004). Conjugated linoleic acids: are the beneficial or detrimental to health? *Prog. Lipid. Res.*

- 43**, 553-587.
- West D.B., Blohm F.Y., Truett A.A. and DeLany J.P. (2000). Conjugated linoleic acid persistently increases total energy expenditure in AKR/J mice without increasing uncoupling protein gene expression. *J. Nutr.* **130**, 2471-2477.
- West D.B., James P.D., Patricia M.C., Fawn B., Alycia A.T. and Joseph S. (1998). Effects of conjugated linoleic acid on body fat and energy metabolism in the mouse. *Am. J. Physiol.* **275**, 667-672.
- Williams C.M. (2000). Dietary fatty acids and human health. *Ann. Zootech.* **49**, 165-180.
- Xu X., Storkson J., Kim S., Sugimoto K., Park Y. and Pariza M.W. (2003). Short-term intake of conjugated linoleic acid inhibits lipoprotein lipase and glucose metabolism but does not enhance lipolysis in mouse adipose tissue. *J. Nutr.* **133**, 663-667.
- Yamasaki M., Ikeda A., Hirao A., Tanaka Y., Miyazaki Y., Rikimaru T., Shimada M., Sugimachi K., Tachibana H. and Yamada K. (2001). Effect of dietary conjugated linoleic acid on the *in vivo* growth of rat hepatoma dRLh-84. *Nutr. Cancer.* **40**, 140-148.
- Yanagita T., Wang Y.M., Nagao K., Ujino Y. and Inoue N. (2005). Conjugated linoleic acid-induced fatty liver can be attenuated by combination with docosahexaenoic acid in C57BL/6N mice. *J. Agric. Food Chem.* **53**, 9629-9633.
- Zanini S.F., Colnago G.L., Pessotti B.M.S., Bastos M.R., Casagrande F.P. and Lima V.R. (2006). Body fat of broiler chickens fed diets with two fat sources and conjugated linoleic acid. *Int. J. Poult. Sci.* **5**, 241-246.
- Zanini S.F., Vicente E., Colnago G.L., Pessotti B.M.S. and Silva M.A. (2008). Manipulation of the fatty acids composition of poultry meat and giblets by dietary inclusion of two oil sources and conjugated linoleic acid. *Arquivo Brasileiro de. Med. Vet. Zootec.* **60**, 1388-1398.
- Zhang H., Guo Y. and Yuan J. (2005). Conjugated linoleic acid enhanced the immune function in broiler chicks. *Br. J. Nutr.* **94**, 746-752.
-